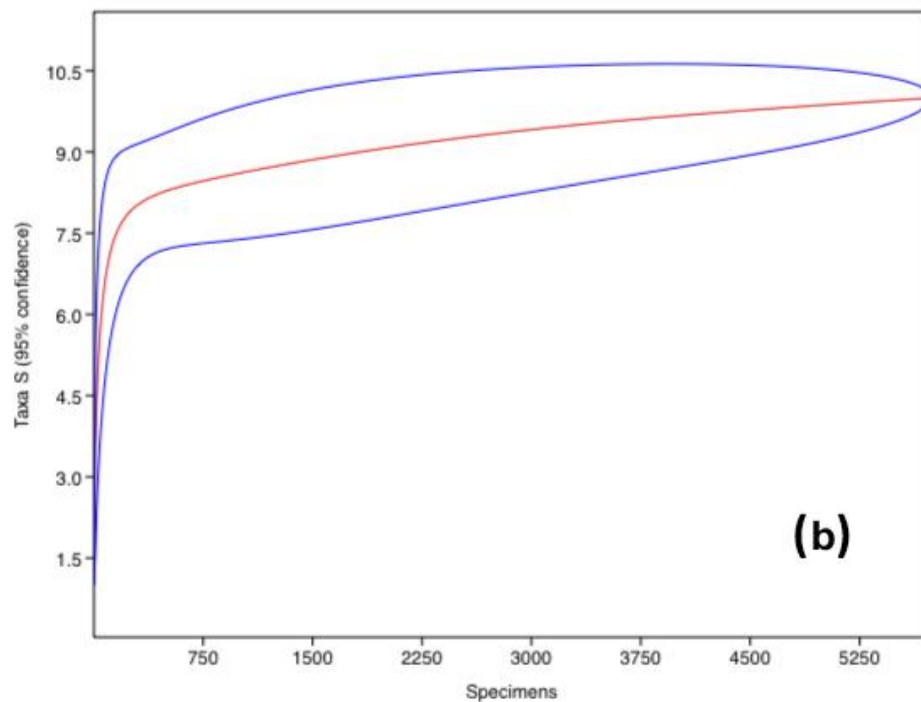
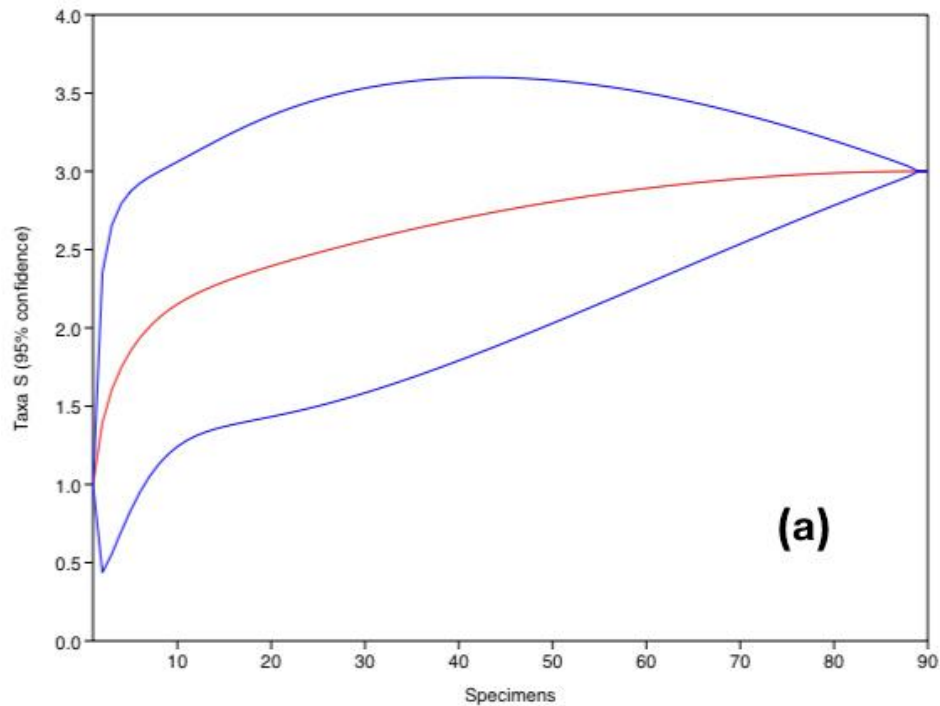
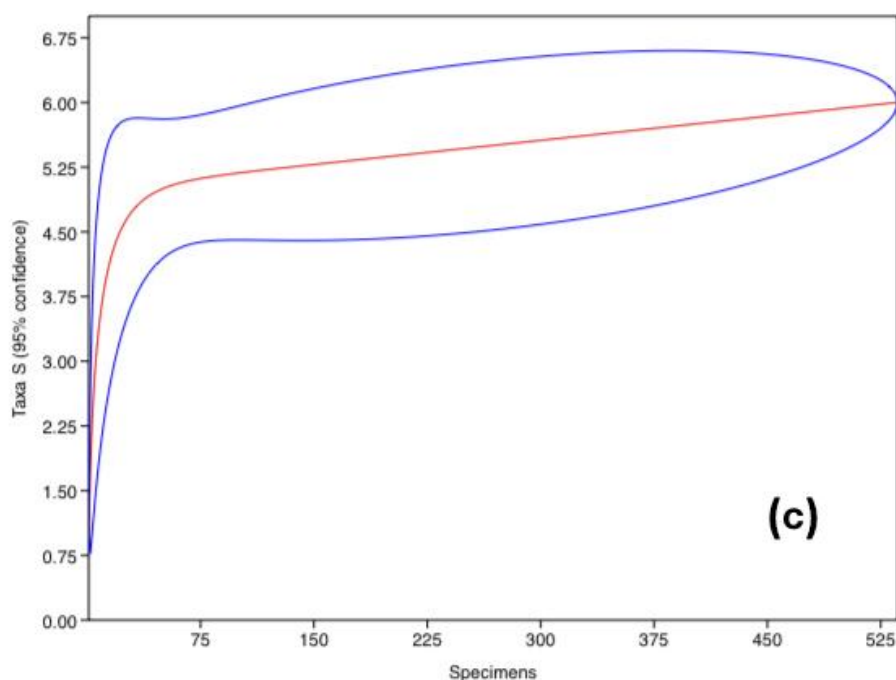


Supplementary material for: Câmara P.E.A.S., Šantl-Temkiv T., Pinto O.H.B., Convey P., Dall'Osto M., Bones F.L.V., Carvalho-Silva M., Knop Henriques D. & Rosa L.R. 2025. DNA metabarcoding of non-fungal eukaryotic diversity in air and snow of Livingston Island, South Shetland Islands, Antarctica. *Polar Research* 44. Correspondence: Paulo E.A.S. Câmara, Departamento de Botânica, Universidade de Brasília, Brasília, Campus Universitário Darcy Ribeiro, Asa Norte, Brazil, 70910-900. E-mail: paducamara@gmail.com.





‘Supplementary Fig. S1. Rarefaction curves based on taxa profile (97% similarity cutoff) from (a) air, (b) Snow1 and (c) Snow2 samples. Blue lines represent 95% confidence limits inferred using bootstrap values calculated from 1000 iterations using PAST, version 1.90.

Methodological details

The methods used here were the same as those used and cited by Rosa et al. (2020).

DNA extraction

DNeasy PowerSoil Kit (Qiagen, USA) following the manufacturer’s instructions. DNA quality was analysed by agarose gel electrophoresis (1% agarose in 1 X Trisborate-EDTA) and then quantified using the Quanti-iT™ Pico Green dsDNA Assay (Invitrogen). Extracted DNA was used as template for generating PCR amplicons. PCR amplicons were generated using the universal primers ITS3 and ITS4 and were sequenced by high-throughput sequencing at Macrogen Inc. (South Korea) on an Illumina MiSeq sequencer, using the MiSeq Reagent Kit version 3 (600-cycle) following the manufacturer’s protocol.

Data analyses and taxa identification

Quality analysis was carried out using BBDuk version 38.87 in BBmap software (Bushnell 2014) with the following parameters: Illumina adapters removing (Illumina artefacts and the PhiX Control v3 Library); ktrim = 1; k = 23; mink = 11; hdist = 1; minlen = 50; tpe; tbo; qtrim = rl; trimq = 20; ftm = 5;maq = 20. The remaining sequences were imported to QIIME2 version 2021.4 ([https:// qiime2.org/](https://qiime2.org/)) for bioinformatics analyses (Bolyen et al.

2019). The qiime2-dada2 plugin was used for filtering, dereplication, turn paired-end fastq files into merged, and remove chimeras, using default parameters (Callahan et al. 2016).

Taxonomic assignments of amplicon sequence variants (ASVs) were determined using the qiime2-feature-classifier (Bokulich et al. 2018) classify-sklearn against different databases, using a sequence similarity threshold of 97%. First, ASVs were classified against the PLANiTS2 database (Banchi et al. 2020). After this step, ASVs that remained unclassified were filtered and classify-sklearn classified against the UNITE Eukaryotes ITS database version 8.3 (Abarenkov et al. 2020).

Finally, remaining unclassified ASVs were filtered and aligned against the filtered National Center for Biotechnology Information (NCBI) non-redundant nucleotide sequences (nt) database (February 2020) using BLASTn (Camacho et al. 2009) with default parameters; the nt database was filtered with the following keywords: “ITS1”, “ITS2”, “Internal transcribed spacer”, and “internal transcribed spacer”.

Taxonomic assignments were performed using MEGAN6 (Huson et al. 2016). For simplicity we henceforth refer to the assigned ASVs as “taxa”. Venn diagrams were prepared as described by Bardou et al. (2014).

For comparative purposes, we consider reads as a proxy for abundance (Deiner et al. 2017, Hering et al. 2018; Câmara, Carvalho-Silva et al. 2021; Câmara, Convey et al. 2021; Carvalho-Silva et al. 2021; Rosa et al. 2021; Câmara et al. 2022).

Diversity analyses

Rarefaction calculations were carried out using the rarefaction analysis command in the software PAST 4.03 (Hammer et al. 2001). Venn diagrams were prepared as described by Bardou et al. (2014).

Supplementary Table S1. Raw data and statistics. All databases were consulted in February 2020.

| DNA reads | Total | Database consulted | | |
|---------------------------------------|---------|-----------------------|--------------------|-------------------|
| | | PLANiTS2 ^a | UNITE ^b | NCBI ^c |
| Number of raw reads | 751 670 | | | |
| Reads after filtering | 439 168 | | | |
| Reads from fungi (Rosa et al. 2020) | 430 748 | | | |
| Reads from non-fungal organisms | 8420 | | | |
| Unassigned reads | 2054 | | | |
| Known reads from non-fungal organisms | 6366 | | | |
| Reads from air sample | 90 | 69 | 21 | 0 |
| Reads from snow samples | 6276 | 5480 | 238 | 46 |

^a <https://github.com/apallavicini/PLANiTS>. ^b <https://unite.ut.ee/>. ^c National Center for Biotechnology Information, <https://blast.ncbi.nlm.nih.gov>.

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